# Chapter 1 Keratin: An Introduction



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**Abstract** What is keratin? And why to use the keratin? Well known that protein is a part of every cell in living organism's body which plays many different roles to keep living things alive and healthy. The importance of protein for the growth and repair of muscles, bones, skin, tendons, ligaments, hair, eyes, and other tissues is proven since a very long time. Proteins also exist in the form of enzymes and hormones needed for metabolism, digestion, and other important processes. Natural proteins are purified from *natural* sources. Keratin is among the most copious proteins found associated with the body of reptiles, birds, and mammals. It is a structural constituent of nail, wool, feathers, and hoofs which offers strength to body and muscles. Nowadays, the keratin-rich waste biomass produced from poultry and meat industry imposes serious threat to environment and living beings. We need to explore various techniques and methods for the extractions and use of keratin from waste biomass. From the industrial point of view, keratin is a useful product in the medical, pharmaceutical, cosmetic, and biotechnological industries. Materials obtained from keratin may be converted into porous foam of different sponges, shapes, coatings, mats, microfibers, gels, and materials of high molecular weight. In this chapter, we briefly describe the various sources, properties, and structures of keratin.

**Keywords** Protein  $\cdot$  Waste  $\cdot$  Biomass  $\cdot$  Medical  $\cdot$  Pharmaceutical  $\cdot$  Application Extraction

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### **1** Introduction

Growing need for sustainable and safe bio-based materials due to rising environmental concern has forced the use of available natural by-products as a substitution. By-products from different animal sources are recently being used for beneficial purposes such as drug delivery, medicines, cosmetics, and bioplastic.

Keratin is among the most abundant structural proteins (Coulombe and Omary 2002), and in animals together with collagen, it is the most important biopolymer (McKittrick et al. 2012). Keratinous materials, formed by specifically organized keratinized cells filled with mainly fibrous proteins (keratins), are natural polymeric composites that exhibit polypeptide chain structure, filament-matrix structure lamellar structure, and sandwich structure ranging from nanoscale to centimeter scale. Keratin is among the most copious proteins found associated with the body of reptiles, birds, and mammals. It is a structural constituent of nail, wool, feathers, and hoofs which offers strength to body and muscles (Reichl et al. 2011). They serve several functions, such as for predation and as armor, protection, and defense. Therefore, a thorough understanding of the relationships between the units that make up a functional keratinous material would expectantly provide useful knowledge in designing new materials. Keratin is chiefly found in epithelial cells in higher vertebrates (Korniłłowicz-Kowalska and Bohacz 2011). Keratins have high strength, stiffness, and insolubility in polar as well as nonpolar solvents. The stabilization is the result of intramolecular and intermolecular disulfide crosslinks, hydrogen bonding, and its crystallinity. These properties will differentiate it from other fibrous proteins like myofibrillar and collagen protein (Schrooyen et al. 2000). With increasing urbanization, food industries particularly the wool industry, slaughterhouse, and meat market produce million tons of keratin-containing biomass. The major producers including USA, Brazil, and China, report for more than 40 million tons annually. These proteins compose keratin by-products containing 15-18% nitrogen, 2-5% sulfur, 1.27% fat, 3.20% mineral elements, and 90% of proteins (Kunert 2000; Sangali and Brandelli 2000; Gessesse et al. 2003).

From past several years, keratin has been extracted from various sources such as horn, hoof, hair, beaks, shells, toenails, claws, fingernails, and feathers (Sharma and Gupta 2016; Sharma et al. 2017c). Keratin was used in medicine for the first time by Chinese herbalist Shi-Zen in sixteenth century. Hoffmeier, in 1905, first time extracted keratin from animal hoofs with the help of lime which further was used in making gels (Rouse and Van Dyke 2010). Keratins are cystine-rich proteins associated with intermediate filaments (IFs) which are cytoskeleton element having diameter of 8–10 nm (ARAI et al. 1983; Khosa et al. 2013). It is available in two forms  $\alpha$ - and  $\beta$ -keratins.  $\alpha$ -keratins are copiously found in the soft tissues such as sheep wool, hair, and skin. These are rich in cystine.  $\beta$ -keratins are present in hard tissue protein of nails, fish scales, bird feathers, and others. They are rich in glycine and alanine, poor in cystine, hydroxyproline, and proline (Gupta et al. 2012). Keratins are very much stable and insoluble in most of organic solvents. The presence of cystine

in ample amount has made the keratin more susceptible to hydrolytic and oxidation reactions (Schrooyen et al. 2000; Barone et al. 2006b; Endo et al. 2008).

Nowadays, a large amount of keratin by-products are unused which is probable hazard to the environment (Cavello et al. 2012; Park et al. 2013). Keratin waste is classified into three categories in regulation (EC) 1774/2002 of the European Parliament and Council of 3rd October 2002 laying down health rules concerning animal by-products which are not intended for human consumption but does not spread diseases to humans or animals (Korniłłowicz-Kowalska and Bohacz 2011). In the form of solid biomass, keratin is less prone to enzymatic hydrolysis due to high cross-linking by hydrogen bonding, disulfide bonds, and hydrophobic interactions (Korniłłowicz-Kowalska and Bohacz 2011).

Thus, use of keratin as a biopolymer requires its extraction from the biomass. In recent years, several attempts have been made for the extraction of keratin using chemical, mechanical, and enzymatic methods (Korol 2012; Jeong et al. 2010; Chaudhari et al. 2013; Fang et al. 2013). There are number of ways for extraction of keratin from the waste biomass including acidic hydrolysis (Breinl and Baudisch 1907; Earland and Knight 1956), alkaline hydrolysis (Tsuda and Nomura 2014; Song et al. 2013; Poole et al. 2008), enzymatic hydrolysis (Eslahi et al. 2013), ionic liquid hydrolysis (Idris et al. 2014; Wang and Cao 2012), and alkaline–enzymatic hydrolysis (Yin et al. 2007). Acidic hydrolysis provides very severe conditions which can destroy some useful amino acids during hydrolysis. Conversely, enzymatic hydrolysis provides less species alteration but with a slower process and is more expensive (Staroń et al. 2014), which makes its commercial use more difficult. On the other hand, ionic liquids are too costly and protein recovery is very low (Cevasco and Chiappe 2014) to be used for industrial purpose. Hence, research into simple, cheap, environmentally sustainable, and industrial applicable method to extract keratin seems justifiable.

Keratin is a useful product in the medical, pharmaceutical, cosmetic, and biotechnological industry. Materials obtained from keratin may be converted into porous foam of different sponges, shapes, coatings, mats, microfibers, gels, and materials of high molecular weight. Keratin is attracting the attention of the researchers due to its abundance. Keratin biomaterial is applied in the development of wound healing gels, tissue engineering, drug delivery, trauma and medical devices, biomedical, and cosmetic applications. One of the impending applications of purified keratin is to produce biomaterials in regeneration and tissue repair (Alsarra 2009; Ramshaw et al. 2009; Natarajan et al. 2012; Ramadass et al. 2013; Kumaran et al. 2016).

These are the polymers formed by various amino acids capable of promoting intraand intermolecular bonds, allowing the resultant materials to have a large variation in their functional properties (Gupta and Perumal 2013). Feather keratin is a potential source of abundant, inexpensive, eco-friendly, and commercial biomaterial (Poole et al. 2009; Shi et al. 2014). Keratin word first emerged around 1850 and it illustrates the material which constitutes hard tissues such as animal horns and hoofs. The name was taken from the word "kera" which means horn. Keratin is an insoluble, highly stable structure, small proteins, and uniform in size. Feather keratin has a molecular weight of about 10 kDa (Fraser et al. 1972; ARAI et al. 1983; Ullah et al. 2011; Kamarudin et al. 2017). It is composed of  $\alpha$ -helix,  $\beta$ -sheet structures as discussed in further chapters. Also, the internal structure of every keratin has  $\alpha$ -helices and  $\beta$ sheets that support the protein. The elastic nature of keratin fiber is due to the interplay between  $\alpha$ -helices and  $\beta$ -sheet configuration of the protein. Feathers consist of 50% of each fiber and quill by weight (Reddy and Yang 2007a). In a feather, fiber has a larger percentage of  $\alpha$ -helix (41%) as compared to  $\beta$ -sheet (38%) and quill fraction consists of more  $\beta$ -sheet (50%) structure than  $\alpha$ -helix (21%) (Barone et al. 2006a; Schmidt and Jayasundera 2004; Wallenberger and Weston 2003; Fraser et al. 1972). According to a previous study (Sun et al. 2009a), feather has 9.38%  $\alpha$ -helix, 47.19%  $\beta$ -sheet, 32.25%  $\beta$ -turn, and 11.18% in random.

Keratin has about 7% cystine, which forms S–S bonds with other cystine molecules (ARAI et al. 1983) and forms cysteine by disulfide bridges. The presence of disulfide, hydrophobic, and hydrogen bond (Onifade et al. 1998; ARAI et al. 1983; Ullah et al. 2011; Cardamone 2010; Bulaj 2005) in keratin provides it strength, mechanical stability, rigidity, and resistance to degradation by proteolytic enzymes such as trypsin, pepsin, and papain (Yamamura et al. 2002; Agrahari and Wadhwa 2010; Paul et al. 2013) to keratin in the solid state. However, these cross-links are a hindrance to processing in the melt state (Barone et al. 2006b). The presence of reactive functional groups, especially peptide backbone, disulfide (–S–S) bridges, amino (–NH<sub>2</sub>), and carboxylic acid (–COOH), makes it chemically reactive under favorable reaction conditions. During controlled reduction, protonation of keratin occurs. Thus, the keratin protein attains positive surface charge and becomes pseudo-cationic biopolymer.

Keratin is insoluble in polar and nonpolar solvents and has very low chemical reactivity. At low pH, high temperature and in presence of reducing agents the solubility of the keratin are increased. The biodegradability and nontoxic nature of keratin make it versatile biopolymer which can be modified and extended in various forms such as films, gel, beads, and nano/microparticles. The modified keratin has plenty of applications in food sciences, green chemistry, cosmetic industries, and pharmaceuticals.

### 2 Chemical Composition and Occurrence of Keratin

Keratin is the most important component of hair, wool, nails, hooves, claws, scales, horn, beaks, and feathers as shown in Table 1. These are least affected by chemical and physical environmental factors (Teresa and Justyna 2011). The keratin extracted is with 90–100 amino acids and 10.2–10.4 kDa molecular weight (Kamarudin et al. 2017; Barone and Schmidt 2005). Chemical structure of keratin showed  $\alpha$ -helix,  $\beta$ -helix, or  $\beta$ -pleated sheet (Fraser et al. 1972; Lee and Baden 1975). Keratin has a high amount of cystine residues (7–15 mol% of amino acids) as compared to other which help to make intermolecular cross-links (Rouse and Van Dyke 2010). The amount of cysteine residues depends on the keratin source, which varies from 7% in feather to 15% in wool keratin (Fraser et al. 1972; ARAI et al. 1983).

<b>Table 1</b> Distribution of $\alpha$ - and $\beta$ -keratins	Types of keratin	Source organ
	α-Keratin	Wool, quills, hair, horns, fingernails, hooves; stratum corneum
	β-Keratin	Feathers, avian beaks and claws, reptilian claws and scales
		$\alpha$ - and $\beta$ -keratin reptilia epidermis, pangolin scales

The high cystine content is the unusual characteristic of keratin which differentiates it from other structural proteins like elastin and collagen. Main amino acids present in keratin are cystine, proline, serine, and glycine (Fraser et al. 1972; Fraser and Parry 2003). In another study,  $\gamma$ -keratin was extracted which is nonstructural and associated with  $\alpha$ - and  $\beta$ -sheets (Fraser et al. 1972; Hill et al. 2010). The sulfur content ratio plays an important role in keratin physical properties. Some researchers based upon sulfur content classified keratin as soft and hard forms (Rizvi and Khan 2008; Zoccola et al. 2009). Soft keratin has lower cystine content, week cross-linking, and smaller resistant to other chemicals found in the hair core and outer layer of epidermis (Fraser et al. 1972).

Hard keratin found in mammalian epidermal appendages, such as hairs, nails, horns, and in avian or reptilian tissues. In recent years, equine hoof (Douglas et al. 1996), bovine hoof, wool (Feughelman and Robinson 1971), and especially the sheep horn (Tombolato et al. 2010) are the attractive sources for keratin extraction.

## 3 Keratin Sources

Keratin biomass is derived from living organisms or from their body parts after death. The major livestock's of keratin includes sheepskins, goatskins, cattle hides, feathers, hairs, and buffalo hides as shown in Fig. 1. Skin and its appendages such as feathers, wool, nails, hooves, hair, scales, and stratum corneum are the richest sources of keratin (Kim 2007). It can be extracted from animal horns and hooves, wool, feathers, and human hairs (Fig. 1). Food industry produces million tons of keratin biomass. About 80% of human hair is formed of keratin only (Kaplin 1982; Wagner and Joekes 2005). It provides flexibility, strength, durability, and functionality to the hair in the form of different conformations (Velasco et al. 2009). Keratinous materials based on  $\alpha$ - and  $\beta$ -keratins are discussed in Table 2.

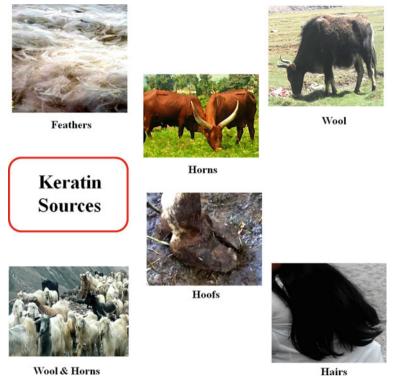


Fig. 1 Main sources of keratin biomass

Keratinous materials based on $\alpha$ -keratin	Keratinous materials based on $\beta$ -keratin	Keratinous materials based on $\alpha$ - and $\beta$ -keratins
Stratum corneum	Feathers	Reptilian epidermis
Wool and hair	Beaks	Hard and soft epidermis of Testudines
Quills	Claws	Pangolin scales
Horns		
Hooves		
Nails		
Whale baleen		
Hagfish slime threads		
Whelk egg capsules		

**Table 2** Keratinous material based on the presence of  $\alpha$ - and  $\beta$ -keratins

#### **4** Applications of the Keratin Protein

Keratin biomass is hydrolyzed by alkali, acid, or enzyme to extract keratin. The extracted keratin has various applications in various industries such as cosmetic, biomedical, and pharmaceutical industries. Furthermore, it does not have harmful effects and thus can be used for variety of cosmetics such as creams, shampoos, hair conditioners, and biomedical products. They have been used as a treatment of skin and human hair as reported before (Weigmann et al. 1990; Innoe 1992). Its existence in the hair cuticle and stratum corneum helps in preserving skin moisture while interacting with cosmetics. Its combination with other natural polymers such as chitosan, collagen, and silk fibroin was used as a component for cosmetic products (Sionkowska 2015). Keratin with high molecular weight is mostly used for skin care applications due to its individuality like film forming and hydrophilic. Keratin film or coating on skin provides smooth and soft sensation. Keratin-associated proteins from different sources were developed and applied as microscaffolds in medicine and cosmetics (Lipkowski et al. 2009). Proteins are useful ingredients for healthy skin and hair. There are studies which reported the role and efficacy of using protein in cosmetics; proteins can be obtained from simple and conjugated proteins (Secchi 2008). There are also studies which described the keratin derivatives and cationizing agent in the various cosmetics having specific functional groups (Matsunaga et al. 1983). Figure 2 shows the hair treatment cream formed using keratin extracted from chicken feathers. Other applications from different biomasses are discussed in Table 3.

Because the presence of high cross-linking by cystine formulation of micro- and nanoparticle from feathers is difficult, some researchers have prepared micro- and nanoparticles successfully from feather keratin. Keratin was converted into useful microparticles by treatment with ionic liquid, 1-butyl-3-methylimidazoliumchloride (Sun et al. 2009b). Treated feathers have low surface area but have higher ion sorption capacity than untreated feathers due to their hydrophilic nature. (Xu et al. 2014b) developed nanoparticles (50-130 nm) from feather keratin which showed good biocompatibility and stability essential for controlled drug release. Here, the chicken feather keratin nanoparticles were developed and used as a haemostatic agent which resulted in decrease in the bleeding time and blood loss in tail amputation and liver scratch rat models (Wang et al. 2016). In another study, quail feathers keratin incorporated into silver nanoparticles and formed the nanofibrous scaffold which gave 99.9% and 98% of the antibacterial activity against Gram-negative (Escherichia coli) and Gram-positive (Staphylococcus aureus) bacteria, respectively. Thus, it can be used for biomedical applications (Khajavi et al. 2016). As chitosan has good properties of biodegradation and biocompatibility, when they are mixed with keratin nanoparticles, they form scaffold. The biodegradation and protein adsorption of the scaffold had increased, and it was noncytotoxic to human osteoblastic cells. Thus, this scaffold can work as biomimetic substrate for bone tissue engineering applications (Saravanan et al. 2013). In one study, keratin nanopowder from chicken feather was produced by electrospraying which has a small particle size and has less crystallinity



Fig. 2 Hair treatment cream made by using keratin extracted from chicken feathers

than raw keratin (Rad et al. 2012). Water stable nanoparticles were developed from feather keratin. Figure 3 shows the keratin nanoparticles extracted from chicken feathers. Nanoparticles can be a good veterinary diagnostic, and it can penetrate

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Sr.	Sources	Industrial application(s)	Reference(s)
no.	Sources	industrial application(s)	Kelefence(s)
1	Human hairs	Medicinal use	Zheng et al. (2005)
2	Hoof and horn's	Preparation of firefighting composition	Datta (1993)
3	Human hair and wool	To explore structural and biological properties of self-assembled keratins	Xu et al. (2014b)
4	Feathers	Development of protein fibers and 2D and 3D scaffolds for tissue engineering	Xu et al. (2014a), Rouse and Van Dyke (2010)
5	Chicken feathers	Keratin film for drug delivery system	Poole et al. (2009), Yin et al. (2013)
		Regenerated fibers	Xu et al. (2014a)
		Micro- and nanoparticles	Sun et al. (2009b)
		Graphene oxide and its derivative in biomaterials	Amieva et al. (2014)
		As a diet supplement for feeding ruminants	Coward-Kelly et al. (2006), Dalev (1994), Dalev et al. (1996), Dalev et al. (1997)
		Microporous material used as electrode material	Zhan and Wool (2011)
		Thermoplastic films	Reddy et al. (2013), Jin et al. (2011)
		Waste management using microorganisms for degradation	Vasileva-Tonkova et al. (2009), Syed et al. (2009), Grazziotin et al. (2006)
		Leather processing	Sastry et al. (1986), Sehgal et al. (1987), Karthikeyan et al. (2007)
		Handspun yarn	Reddy and Yang (2007b)
		Textile yarns	Reddy et al. (2014a, b), Yang and Reddy (2013)
		Keratinases in detergents formulation	Balakumar et al. (2013), Manivasagan et al. (2014), Rai et al. (2009)
		Flame retardant	Wang et al. (2014)
		Bio-composites or composite fabrication	Flores-Hernández et al. (2014), Spiridon et al. (2012), Huda and Yang (2008, 2009)
		Biofertilizer	Ichida et al. (2001), Kornillowicz-Kowalska and Bohacz (2010), Gurav and Jadhav (2013), Hadas and Kautsky (1994), Gousterova et al. (2012)
			(continue

 Table 3 Applications from different keratin biomasses

(continued)

Sr. no.	Sources	Industrial application(s)	Reference(s)
		Nanoparticle and microparticles for pharmaceutical application	Xu et al. (2014b), Sundaram et al. (2015), Yu et al. (2014), Sharma et al. (2017a, b)
		Keratin and graphene oxides within biomaterials	Amieva et al. (2014)
		Cement-bonded composites	Acda (2010)
		Keratin hydrogel	Wang et al. (2017), Barati et al. (2017), Priyaah et al. (2017)
		Tissue regenerative applications	Li et al. (2013), Saravanan et al. (2013), Kumar et al. (2017)
		Paper production	Tesfaye et al. (2017)
		Bioplastic	Sharma et al. (2018), Ramakrishnan et al. (2018)

Table 3 (continued)

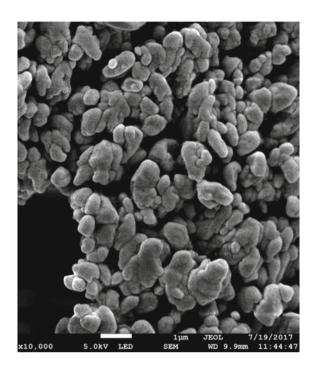
easily into cells and organs due to their nano size. They are good for the drug delivery as compared to synthetic polymer and carbohydrates. Xu et al. (2014b) found that keratin nanoparticles have supportive function for the cell growth and stable in physiological environment for up to 7 days. The keratin nanoparticle shows anticancer properties. When keratin nanoparticles combine with chlorin e6, it resulted in greater cell death percentage (90%) as compared to free chlorin e6 on osteosarcoma (U2OS) and glioblastoma (U87) cells lines, and thus keratin nanoparticles are effective and promising delivery vehicles for photodynamic therapy applications (Aluigi et al. 2016). In one study, keratin-based drug-loaded nanoparticles were made and it was showing pH and glutathione dual-responsive behavior. Thus, from the results of the study conducted, it was concluded that keratin-based drug carriers had potential for drug delivery and cancer therapy in clinical medicine (Li et al. 2017).

Biocompatible composites were developed which can be used for highperformance dressing, to treat chronic ulcerous infected wounds by the combination of cellulose and keratin with the silver nanoparticles (Tran et al. 2016). Hair keratin nanoparticle had faster clotting time, and it significantly reduces blood loss and coagulation time which forms the viscosity gel on wound; hence, it has a great potential for haemostatic application (Luo et al. 2016). The soluble keratin would have applications in tissue regeneration, cell seeding, wound healing, and drug delivery (Yamini Satyawali 2013). Soluble keratin can be used to make 2D and 3D scaffolds, and protein fibers which further utilized for tissue engineering (Xu et al. 2014b). Due to self-congregation and polymerization property of keratin proteins has led to work as scaffolds for tissue engineering (Rouse and Van Dyke 2010).

When the partially oxidized keratin is added into water, it forms the hydrogel which can be used as an absorbent material, as a therapeutic for skin. The hydrogel can be used for the implantation. The keratin can be incorporated into films which are

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Fig. 3 Keratin nanoparticles extracted from chicken feathers



nonwoven, and these two materials are suitable for use in tissue engineering scaffolds (Blanchard et al. 2002). Keratin biomaterials have the potential to interact with cells and tissues but the composition, structure, and cell-instructive characteristics are not clear. Burnett et al. (2013) made keratin-based biomaterial, demonstrate self-assembly of cross-linked hydrogels, investigate a cell-specific interaction, and find the utilization in drug and cell delivery, tissue engineering, regenerative medicine, and trauma. The huge amount of waste biomass generated by animals as well as food industries can be used as raw material for the production of keratin at industrial level. The keratin waste biomass management by reconversion into industrially used products will save the ecosystem from large amount of sludge and boost up industries such as pharmaceutical and cosmetic industry economically.

# 5 Conclusion

To develop the strategies for efficient extraction of keratin from poultry biomass will prove to be very beneficial for sustainable management of huge waste. Researchers are working to develop various chemical, biological, and physical methods individually as well as in combined form for keratin extraction. The insoluble protein has many advantages in biomedical industry to develop products of pharmaceutical use, in tissue engineering as well as in agriculture industry too. Recent advances in the sustainable management of poultry waste biomass, extraction techniques used by various researchers, and numerous applications of the extractions have been discussed in the subsequent chapters of this book.

### References

- Acda MN (2010) Waste chicken feather as reinforcement in cement-bonded composites. Philippine J Sci 139(2):161–166
- Agrahari S, Wadhwa N (2010) Degradation of chicken feather a poultry waste product by keratinolytic bacteria isolated from dumping site at Ghazipur poultry processing plant. Int J Poult Sci 9(5):482–489
- Alsarra IA (2009) Chitosan topical gel formulation in the management of burn wounds. Int J Biol Macromol 45(1):16–21
- Aluigi A, Sotgiu G, Ferroni C, Duchi S, Lucarelli E, Martini C, Posati T, Guerrini A, Ballestri M, Corticelli F (2016) Chlorin e6 keratin nanoparticles for photodynamic anticancer therapy. RSC Adv 6(40):33910–33918
- Amieva EJ-C, Fuentes-Ramirez R, Martinez-Hernandez A, Millan-Chiu B, Lopez-Marin LM, Castaño V, Velasco-Santos C (2014) Graphene oxide and reduced graphene oxide modification with polypeptide chains from chicken feather keratin. J Alloy Compd 643:S137–S143
- Arai KM, Takahashi R, Yokote Y, Akahane K (1983) Amino-acid sequence of feather keratin from fowl. Eur J Biochem 132(3):501–507
- Balakumar S, Mahesh N, Arunkumar M, Sivakumar R, Hemambujavalli V (2013) Optimization of keratinase production by keratinolytic organisms under submerged fermentation. Optimization 5(3):1294–1300
- Barati D, Kader S, Pajoum Shariati SR, Moeinzadeh S, Sawyer RH, Jabbari E (2017) Synthesis and characterization of photo-cross-linkable keratin hydrogels for stem cell encapsulation. Biomacromolecules 18(2):398–412
- Barone JR, Dangaran K, Schmidt WF (2006a) Blends of cysteine-containing proteins. J Agric Food Chem 54(15):5393–5399
- Barone JR, Schmidt WF (2005) Polyethylene reinforced with keratin fibers obtained from chicken feathers. Compos Sci Technol 65(2):173–181
- Barone JR, Schmidt WF, Gregoire N (2006b) Extrusion of feather keratin. J Appl Polym Sci 100(2):1432–1442
- Blanchard CR, Van Dyke ME, Timmons SF, Siller-Jackson AJ, Smith RA (2002) Non-woven keratin cell scaffold. Google Patents
- Breinl F, Baudisch O (1907) The oxidative breaking up of keratin through treatment with hydrogen peroxide. Z Physiol Chem 52:158–169
- Bulaj G (2005) Formation of disulfide bonds in proteins and peptides. Biotechnol Adv 23(1):87-92
- Burnett LR, Rahmany MB, Richter JR, Aboushwareb TA, Eberli D, Ward CL, Orlando G, Hantgan RR, Van Dyke ME (2013) Hemostatic properties and the role of cell receptor recognition in human hair keratin protein hydrogels. Biomaterials 34(11):2632–2640
- Cardamone JM (2010) Investigating the microstructure of keratin extracted from wool: Peptide sequence (MALDI-TOF/TOF) and protein conformation (FTIR). J Mol Struct 969(1):97–105
- Cavello I, Cavalitto S, Hours R (2012) Biodegradation of a keratin waste and the concomitant production of detergent stable serine proteases from *Paecilomyces lilacinus*. Appl Biochem Biotechnol 167(5):945–958
- Cevasco G, Chiappe C (2014) Are ionic liquids a proper solution to current environmental challenges? Green Chem 16(5):2375–2385

- Chaudhari PN, Chincholkar SB, Chaudhari BL (2013) Biodegradation of feather keratin with a PEGylated protease of chryseobacterium gleum. Process Biochem 48(12):1952–1963. https://doi.org/10.1016/j.procbio.2013.09.011
- Coulombe PA, Omary MB (2002) 'Hard'and 'soft' principles defining the structure, function and regulation of keratin intermediate filaments. Curr Opin Cell Biol 14(1):110–122
- Coward-Kelly G, Agbogbo FK, Holtzapple MT (2006) Lime treatment of keratinous materials for the generation of highly digestible animal feed: 2. Animal hair. Bioresour Technol 97(11):1344–1352
- Dalev P, Ivanov I, Liubomirova A (1997) Enzymic modification of feather keratin hydrolysates with lysine aimed at increasing the biological value. J Sci Food Agric 73(2):242–244
- Dalev P, Ljubomirova A, Simeonova L, Ivanov I (1996) Protein hydrolysates from waste feathers for feed and their enrichment with lysine trough enzyme catalyzed covalent binding. In: Mededelingen-faculteit landbouwkundige en toegepaste biologische wetenschappen, vol 61, pp 1641–1644
- Dalev PG (1994) Utilisation of waste feathers from poultry slaughter for production of a protein concentrate. Biores Technol 48(3):265–267
- Datta M (1993) Role of keratin in fire fighting. J Ind Leath Technol Assoc 43:297-299
- Douglas J, Mittal C, Thomason J, Jofriet J (1996) The modulus of elasticity of equine hoof wall: implications for the mechanical function of the hoof. J Exp Biol 199(8):1829–1836
- Earland C, Knight C (1956) Studies on the structure of keratin II. The amino acid content of fractions isolated from oxidized wool. Biochimica et Biophysica Acta 22(3):405–411
- Endo R, Kamei K, Iida I, Kawahara Y (2008) Dimensional stability of waterlogged wood treated with hydrolyzed feather keratin. J Archaeol Sci 35(5):1240–1246
- Eslahi N, Dadashian F, Nejad NH (2013) An investigation on keratin extraction from wool and feather waste by enzymatic hydrolysis. Prep Biochem Biotechnol 43(7):624–648
- Fang Z, Zhang J, Liu BH, Du GC, Chen J (2013) Biodegradation of wool waste and keratinase production in scale-up fermenter with different strategies by *Stenotrophomonas maltophilia* BBE11-1. Bioresource Technol 140:286–291. https://doi.org/10.1016/j.biortech.2013.04.091
- Feughelman M, Robinson M (1971) Some mechanical properties of wool fibers in the "Hookean" region from zero to 100% relative humidity. Text Res J 41(6):469–474
- Flores-Hernández CG, Colín-Cruz A, Velasco-Santos C, Castaño VM, Rivera-Armenta JL, Almendarez-Camarillo A, García-Casillas PE, Martínez-Hernández AL (2014) All green composites from fully renewable biopolymers: chitosan-starch reinforced with keratin from feathers. Polymers 6(3):686–705
- Fraser R, MacRae T, Rogers GE (1972) Keratins: their composition, structure, and biosynthesis. Thomas, Charles C
- Fraser RDB, Parry DAD (2003) Macrofibril assembly in trichocyte (hard alpha-) keratins. J Struct Biol 142(2):319–325. https://doi.org/10.1016/S1047-8477(03)00027-3
- Gessesse A, Hatti-Kaul R, Gashe BA, Mattiasson B (2003) Novel alkaline proteases from alkaliphilic bacteria grown on chicken feather. Enzyme and Microbial Technology 32(5):519–524
- Gousterova A, Nustorova M, Paskaleva D, Naydenov M, Neshev G, Vasileva-Tonkova E (2012) Assessment of feather hydrolysate from thermophilic actinomycetes for soil amendment and biological control application. Int J Environ Res 6(2):467–474
- Grazziotin A, Pimentel FA, de Jong EV, Brandelli A (2006) Nutritional improvement of feather protein by treatment with microbial keratinase. Anim Feed Sci Tech 126(1–2):135–144. https:// doi.org/10.1016/j.anifeedsci.2005.06.002
- Gupta A, Kamarudin NB, Kee CYG, Yunus RBM (2012) Extraction of keratin protein from chicken feather. J Chem Chem Eng 6(8):732
- Gupta A, Perumal R (2013) Process for extracting keratin. Google Patents
- Gurav RG, Jadhav JP (2013) A novel source of biofertilizer from feather biomass for banana cultivation. Environ Sci Pollut R 20(7):4532–4539. https://doi.org/10.1007/s11356-012-1405-z
- Hadas A, Kautsky L (1994) Feather meal, a semi-slow-release nitrogen fertilizer for organic farming. Fertil Res 38(2):165–170

- Hill P, Brantley H, Van Dyke M (2010) Some properties of keratin biomaterials: kerateines. Biomaterials 31(4):585–593. https://doi.org/10.1016/j.biomaterials.2009.09.076
- Huda S, Yang YQ (2008) Composites from ground chicken quill and polypropylene. Compos Sci Technol 68(3–4):790–798. https://doi.org/10.1016/j.compscitech.2007.08.015
- Huda S, Yang YQ (2009) Feather fiber reinforced light-weight composites with good acoustic properties. J Polym Environ 17(2):131–142. https://doi.org/10.1007/s10924-009-0130-2
- Ichida JM, Krizova L, LeFevre CA, Keener HM, Elwell DL, Burtt EH (2001) Bacterial inoculum enhances keratin degradation and biofilm formation in poultry compost. J Microbiol Meth 47(2):199–208. https://doi.org/10.1016/S0167-7012(01)00302-5
- Idris A, Vijayaraghavan R, Patti A, MacFarlane D (2014) Distillable protic ionic liquids for keratin dissolution and recovery. ACS Sustain Chem Eng 2(7):1888–1894
- Innoe T (1992) Hair cosmetic for protection of skins. Eur Pat Appl, EP 469
- Jeong JH, Lee OM, Jeon YD, Kim JD, Lee NR, Lee CY, Son HJ (2010) Production of keratinolytic enzyme by a newly isolated feather-degrading Stenotrophomonas maltophilia that produces plant growth-promoting activity. Process Biochem 45(10):1738–1745. https://doi.org/10. 1016/j.procbio.2010.07.020
- Jin E, Reddy N, Zhu Z, Yang Y (2011) Graft polymerization of native chicken feathers for thermoplastic applications. J Agr Food Chem 59(5):1729–1738
- Kamarudin NB, Sharma S, Gupta A, Kee CG, Chik SMSBT, Gupta R (2017) Statistical investigation of extraction parameters of keratin from chicken feather using Design-Expert. 3 Biotech 7(2):127
- Kaplin I (1982) Effect of cosmetic treatments on the ultrastructure of hair. Cosmet Toiletries 97(8):22-26
- Karthikeyan R, Balaji S, Sehgal P (2007) Industrial applications of keratins—a review. J Sci Ind Res 66(9):710
- Khajavi R, Rahimi MK, Abbasipour M, Brendjchi AH (2016) Antibacterial nanofibrous scaffolds with lowered cytotoxicity using keratin extracted from quail feathers. J Bioact Compat Polym 31(1):60–71
- Khosa MA, Wu J, Ullah A (2013) Chemical modification, characterization, and application of chicken feathers as novel biosorbents. RSC Adv 3(43):20800–20810
- Kim J-D (2007) Purification and characterization of a keratinase from a feather-degrading fungus, *Aspergillus flavus* strain K-03. Mycobiology 35(4):219–225
- Kornillowicz-Kowalska T, Bohacz J (2010) Dynamics of growth and succession of bacterial and fungal communities during composting of feather waste. Bioresource Technol 101(4):1268–1276. https://doi.org/10.1016/j.biortech.2009.09.053
- Korniłłowicz-Kowalska T, Bohacz J (2011) Biodegradation of keratin waste: theory and practical aspects. Waste Manag 31(8):1689–1701
- Korol J (2012) Polyethylene matrix composites reinforced with keratin fibers obtained from waste chicken feathers. J Biob Mater Bio 6(4):355–360. https://doi.org/10.1166/jbmb.2012.1237
- Kumar SL, Anandhavelu S, Sivaraman J, Swathy M (2017) Modified extraction and characterization of keratin from Indian goat hoof: a biocompatible biomaterial for tissue regenerative applications. Integr Ferroelectr 184(1):41–49
- Kumaran P, Gupta A, Sharma S (2016) Synthesis of wound-healing keratin hydrogels using chicken feathers proteins and its properties. 2016:8. https://doi.org/10.22159/ijpps.2017v9i2.15620
- Kunert J (2000) Physiology of keratinophilic fungi. Revista Iberoamericana de Micologia 1:77-85
- Lee L, Baden H (1975) Chemistry and composition of the keratins. Int J Dermatol 14(3):161–171
- Li J-S, Li Y, Liu X, Zhang J, Zhang Y (2013) Strategy to introduce an hydroxyapatite-keratin nanocomposite into a fibrous membrane for bone tissue engineering. J Mater Chem B 1(4):432–437
- Li Y, Zhi X, Lin J, You X, Yuan J (2017) Preparation and characterization of DOX loaded keratin nanoparticles for pH/GSH dual responsive release. Mater Sci Eng, C 73:189–197
- Lipkowski AW, Gajkowska B, Grabowska A, Kurzepa K (2009) Keratin-associated protein micromaterials for medical and cosmetic applications. Polimery 54(5):386–388

- Luo T, Hao S, Chen X, Wang J, Yang Q, Wang Y, Weng Y, Wei H, Zhou J, Wang B (2016) Development and assessment of kerateine nanoparticles for use as a hemostatic agent. Mater Sci Eng, C 63:352–358
- Manivasagan P, Sivakumar K, Gnanam S, Venkatesan J, Kim S-K (2014) Production, biochemical characterization and detergents application of keratinase from the marine actinobacterium *Actinoalloteichus* sp. MA-32. J Surfactants Deterg 17 (4):669–682
- Matsunaga K, Okumura T, Tsushima R (1983) Hair treatment cosmetics containing cationic keratin derivatives. Google Patents
- McKittrick J, Chen P-Y, Bodde S, Yang W, Novitskaya E, Meyers M (2012) The structure, functions, and mechanical properties of keratin. JOM 64(4):449–468
- Natarajan K, Xie Y, Baer MR, Ross DD (2012) Role of breast cancer resistance protein (BCRP/ABCG2) in cancer drug resistance. Biochem Pharmacol 83(8):1084–1103
- Onifade A, Al-Sane N, Al-Musallam A, Al-Zarban S (1998) A review: potentials for biotechnological applications of keratin-degrading microorganisms and their enzymes for nutritional improvement of feathers and other keratins as livestock feed resources. Biores Technol 66(1):1–11
- Park M, Kim B-S, Shin HK, Park S-J, Kim H-Y (2013) Preparation and characterization of keratin-based biocomposite hydrogels prepared by electron beam irradiation. Mater Sci Eng, C 33(8):5051–5057
- Paul T, Halder SK, Das A, Bera S, Maity C, Mandal A, Das PS, Mohapatra PKD, Pati BR, Mondal KC (2013) Exploitation of chicken feather waste as a plant growth promoting agent using keratinase producing novel isolate Paenibacillus woosongensis TKB2. Biocatal Agric Biotechnol 2(1):50–57
- Poole AJ, Church JS, Huson MG (2008) Environmentally sustainable fibers from regenerated protein. Biomacromolecules 10(1):1–8
- Poole AJ, Church JS, Huson MG (2009) Environmentally sustainable fibers from regenerated protein. Biomacromolecules 10(1):1–8. https://doi.org/10.1021/bm8010648
- Priyaah K, Gupta A, Sharma S (2017) Synthesis of wound-healing keratin hydrogels using chicken feathers proteins and its properties. Int J Pharm Pharm Sci 9(2):171–178
- Rad ZP, Tavanai H, Moradi A (2012) Production of feather keratin nanopowder through electrospraying. J Aerosol Sci 51:49–56
- Rai SK, Konwarh R, Mukherjee AK (2009) Purification, characterization and biotechnological application of an alkaline β-keratinase produced by *Bacillus subtilis* RM-01 in solid-state fermentation using chicken-feather as substrate. Biochem Eng J 45(3):218–225
- Ramadass SK, Perumal S, Jabaris SL, Madhan B (2013) Preparation and evaluation of mesalamine collagen in situ rectal gel: a novel therapeutic approach for treating ulcerative colitis. Eur J Pharm Sci 48(1):104–110
- Ramakrishnan N, Sharma S, Gupta A, Alashwal BY (2018) Keratin based bioplastic film from chicken feathers and its characterization. Int J Biol Macromol
- Ramshaw JA, Peng YY, Glattauer V, Werkmeister JA (2009) Collagens as biomaterials. J Mater Sci Mater Med 20(1):3
- Reddy N, Chen L, Zhang Y, Yang Y (2014a) Reducing environmental pollution of the textile industry using keratin as alternative sizing agent to poly (vinyl alcohol). J Clean Prod 65:561–567
- Reddy N, Jiang QR, Jin EQ, Shi Z, Hou XL, Yang YQ (2013) Bio-thermoplastics from grafted chicken feathers for potential biomedical applications. Colloid Surf B 110:51–58. https://doi.org/ 10.1016/j.colsurfb.2013.04.019
- Reddy N, Shi Z, Temme L, Xu H, Xu L, Hou X, Yang Y (2014b) Development and characterization of thermoplastic films from sorghum distillers dried grains grafted with various methacrylates. J Agr Food Chem 62(11):2406–2411
- Reddy N, Yang Y (2007a) Structure and properties of chicken feather barbs as natural protein fibers. J Polym Environ 15(2):81–87
- Reddy N, Yang YQ (2007b) Structure and properties of chicken feather barbs as natural protein fibers. J Polym Environ 15(2):81–87. https://doi.org/10.1007/s10924-007-0054-7
- Reichl S, Borrelli M, Geerling G (2011) Keratin films for ocular surface reconstruction. Biomaterials 32(13):3375–3386

- Rizvi TZ, Khan MA (2008) Temperature-dependent dielectric properties of slightly hydrated horn keratin. Int J Biol Macromol 42(3):292–297. https://doi.org/10.1016/j.ijbiomac.2008.01.001
- Rouse JG, Van Dyke ME (2010) A review of keratin-based biomaterials for biomedical applications. Materials 3(2):999–1014
- Sangali S, Brandelli A (2000) Feather keratin hydrolysis by a Vibrio sp. strain kr2. J Appl Microbiol 89(5):735–743
- Saravanan S, Sameera D, Moorthi A, Selvamurugan N (2013) Chitosan scaffolds containing chicken feather keratin nanoparticles for bone tissue engineering. Int J Biol Macromol 62:481–486
- Sastry T, Sehgal P, Gupta K, Kumar M (1986) Solubilised keratins as a filler in the retanning of upper leathers. Leather Sci 33:345
- Schmidt W, Jayasundera S (2003) In: Wallenberger F, Weston N (eds) Natural fibers plastics, and composites-recent advances. Kluwer Academic: USA
- Schmidt WF, Jayasundera S. (2004) Microcrystalline Avian Keratin Protein Fibers. In: Wallenberger FT, Weston NE (eds) Natural Fibers, Plastics and Composites. Springer, Boston, MA
- Schrooyen PM, Dijkstra PJ, Oberthür RC, Bantjes A, Feijen J (2000) Partially carboxymethylated feather keratins. 1. Properties in aqueous systems. J Agric Food Chem 48(9):4326–4334
- Secchi G (2008) Role of protein in cosmetics. Clin Dermatol 26(4):321-325
- Sehgal P, Sastry T, Kumar M (1987) Effect of keratin filler in retanning of nappa garment leathers. Leather Sci 34(1)
- Sharma S, Gupta A (2016) Sustainable management of keratin waste biomass: applications and future perspectives. Braz Arch Biol Technol 59
- Sharma S, Gupta A, Chik SMS, Kee CG, Mistry BM, Kim DH, Sharma G (2017a) Characterization of keratin microparticles from feather biomass with potent antioxidant and anticancer activities. Int J Biol Macromol 104:189–196
- Sharma S, Gupta A, Chik SMSBT, Kee CYG, Poddar PK (2017b) Dissolution and characterization of biofunctional keratin particles extracted from chicken feathers. In: IOP conference series: materials science and engineering, vol 1. IOP Publishing, p 012013
- Sharma S, Gupta A, Chik SMST, Kee CYG, Podder PK, Subramaniam M, Thuraisingam J (2017c) Study of different treatment methods on chicken feather biomass. IIUM Eng J 18(2):47–55
- Sharma S, Gupta A, Kumar A, Kee CG, Kamyab H, Saufi SM (2018) An efficient conversion of waste feather keratin into ecofriendly bioplastic film. Clean Technol Environ Policy 1–11
- Shi Z, Reddy N, Hou XL, Yang YQ (2014) Tensile properties of thermoplastic feather films grafted with different methacrylates. ACS Sustain Chem Eng 2(7):1849–1856. https://doi.org/10.1021/ sc500201q
- Sionkowska A (2015) The potential of polymers from natural sources as components of the blends for biomedical and cosmetic applications. Pure Appl Chem 87(11–12):1075–1084
- Song N-B, Jo W-S, Song H-Y, Chung K-S, Won M, Song KB (2013) Effects of plasticizers and nano-clay content on the physical properties of chicken feather protein composite films. Food Hydrocolloids 31(2):340–345
- Spiridon I, Paduraru OM, Rudowski M, Kozlowski M, Darie RN (2012) Assessment of changes due to accelerated weathering of low-density polyethylene/feather composites. Ind Eng Chem Res 51(21):7279–7286. https://doi.org/10.1021/ie300738d
- Staroń P, Banach M, Kowalski Z, Staroń A (2014) Hydrolysis of keratin materials derived from poultry industry. In: Proceedings of ECOpole 8
- Sun P, Liu Z-T, Liu Z-W (2009a) Particles from bird feather: a novel application of an ionic liquid and waste resource. J Hazard Mater 170(2):786–790
- Sun P, Liu ZT, Liu ZW (2009b) Particles from bird feather: a novel application of an ionic liquid and waste resource. J Hazard Mater 170(2–3):786–790. https://doi.org/10.1016/j.jhazmat.2009. 05.034
- Sundaram M, Legadevi R, Banu NA, Gayathri V, Palanisammy A (2015) A study on anti bacterial activity of keratin nanoparticles from chicken feather waste against *Staphylococcus aureus* (Bovine Mastitis Bacteria) and its anti oxidant activity. Eur J Biotechnol Biosci 6:1–5

- Syed DG, Lee JC, Li WJ, Kim CJ, Agasar D (2009) Production, characterization and application of keratinase from *Streptomyces gulbargensis*. Bioresource Technol 100(5):1868–1871. https:// doi.org/10.1016/j.biortech.2008.09.047
- Teresa KK, Justyna B (2011) Biodegradation of keratin waste: theory and practical aspects. Waste Manage 31(8):1689–1701. https://doi.org/10.1016/j.wasman.2011.03.024
- Tesfaye T, Sithole B, Ramjugernath D, Chunilall V (2017) Valorisation of chicken feathers: application in paper production. J Clean Prod 164:1324–1331
- Tombolato L, Novitskaya EE, Chen P-Y, Sheppard FA, McKittrick J (2010) Microstructure, elastic properties and deformation mechanisms of horn keratin. Acta Biomater 6(2):319–330
- Tran CD, Prosenc F, Franko M, Benzi G (2016) One-pot synthesis of biocompatible silver nanoparticle composites from cellulose and keratin: characterization and antimicrobial activity. ACS Appl Mater Interfaces
- Tsuda Y, Nomura Y (2014) Properties of alkaline-hydrolyzed waterfowl feather keratin. Anim Sci J 85(2):180–185
- Ullah A, Vasanthan T, Bressler D, Elias AL, Wu J (2011) Bioplastics from feather quill. Biomacromolecules 12(10):3826–3832
- Vasileva-Tonkova E, Gousterova A, Neshev G (2009) Ecologically safe method for improved feather wastes biodegradation. Int Biodeter Biodegr 63(8):1008–1012. https://doi.org/10.1016/j.ibiod. 2009.07.003
- Velasco MVR, Dias TCdS, Freitas AZd, Júnior NDV, Pinto CASdO, Kaneko TM, Baby AR (2009) Hair fiber characteristics and methods to evaluate hair physical and mechanical properties. Braz J Pharm Sci 45(1):153–162
- Wagner RdCC, Joekes I (2005) Hair protein removal by sodium dodecyl sulfate. Colloids Surf, B 41(1):7–14
- Wallenberger FT, Weston N (2003) Natural fibers, plastics and composites. Springer Science & Business Media,
- Wang J, Hao S, Luo T, Cheng Z, Li W, Gao F, Guo T, Gong Y, Wang B (2017) Feather keratin hydrogel for wound repair: preparation, healing effect and biocompatibility evaluation. Colloids Surf, B 149:341–350
- Wang J, Hao S, Luo T, Yang Q, Wang B (2016) Development of feather keratin nanoparticles and investigation of their hemostatic efficacy. Mater Sci Eng, C 68:768–773
- Wang XY, Lu CQ, Chen CX (2014) Effect of chicken-feather protein-based flame retardant on flame retarding performance of cotton fabric. J Appl Polym Sci 131(15). https://doi.org/10.1002/app. 40584
- Wang Y-X, Cao X-J (2012) Extracting keratin from chicken feathers by using a hydrophobic ionic liquid. Process Biochem 47(5):896–899
- Weigmann H, Kamath Y, Ruetsch S (1990) Characterization of surface deposits on human hair fibers. J Soc Cosmet Chem 41:379–390
- Xu H, Cai S, Xu L, Yang Y (2014a) Water-stable three-dimensional ultrafine fibrous scaffolds from keratin for cartilage tissue engineering. Langmuir 30(28):8461–8470
- Xu H, Shi Z, Reddy N, Yang Y (2014b) Intrinsically water-stable keratin nanoparticles and their in vivo biodistribution for targeted delivery. J Agric Food Chem 62(37):9145–9150
- Yamamura S, Morita Y, Hasan Q, Yokoyama K, Tamiya E (2002) Keratin degradation: a cooperative action of two enzymes from *Stenotrophomonas* sp. Biochem Biophys Res Commun 294(5):1138–1143
- Yamini Satyawali SS (2013) Enzymatic electrosynthesis: an overview on the progress in enzymeelectrodes for the production of electricity, fuels and chemicals. J Microb Biochem Technol. https://doi.org/10.4172/1948-5948.s6-007
- Yang YQ, Reddy N (2013) Potential of using plant proteins and chicken feathers for cotton warp sizing. Cellulose 20(4):2163–2174. https://doi.org/10.1007/s10570-013-9956-9
- Yin J, Rastogi S, Terry AE, Popescu C (2007) Self-organization of oligopeptides obtained on dissolution of feather keratins in superheated water. Biomacromolecules 8(3):800–806

- Yin XC, Li FY, He YF, Wang Y, Wang RM (2013) Study on effective extraction of chicken feather keratins and their films for controlling drug release. Biomater Sci-UK 1(5):528–536. https://doi. org/10.1039/c3bm00158j
- Yu D, Cai JY, Liu X, Church JS, Wang L (2014) Novel immobilization of a quaternary ammonium moiety on keratin fibers for medical applications. Int J Biol Macromol 70:236–240
- Zhan M, Wool RP (2011) Mechanical properties of chicken feather fibers. Polym Compos 32(6):937-944
- Zheng Y, Du X, Wang W, Boucher M, Parimoo S, Stenn K (2005) Organogenesis from dissociated cells: generation of mature cycling hair follicles from skin-derived cells. J Invest Dermatol 124(5):867–876
- Zoccola M, Aluigi A, Tonin C (2009) Characterisation of keratin biomass from butchery and wool industry wastes. J Mol Struct 938(1–3):35–40. https://doi.org/10.1016/j.molstruc.2009.08.036